

Academic Council Meeting No. and Date : 4 / June 14, 2022

Agenda Number : 2

Resolution Number : 4.6 & 4.14



Vidya Prasarak Mandal's
B. N. Bandodkar College of
Science (Autonomous), Thane



Syllabus for
Programme : Bachelor of Science

Specific Programme : Microbiology

[S.Y.B.Sc. Microbiology]

**Revised under Autonomy
From academic year 2022 - 2023**

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PREAMBLE

Microbiology is one of the most promising branches of modern biology. It is developing with fast pace and generating interest in many young scientists engaged in interdisciplinary research.

With the objective of exposing preliminary learners to many related field and applications of basic microbiology concepts the attempt has been made to build the syllabus of S.Y. B.Sc.Microbiology.In addition to imparting knowledge of basic skill set, deliberate efforts are being made to help learners apply the scientific knowledge in his/her day-to-day life, by engaging the learners in interesting laboratory experimentation with real-time experience.

Continuing the Choice Based Credit System (CBCS) implemented by the esteemed University from the academic year 2017-2018, the existing syllabus of S.Y.B.Sc.Microbiology is restructured according to the CBCS pattern for its implementation from 2022-23 under the autonomous status of VPM's B. N. Bandodkar College of Science. While earlier revision of the syllabus took care of balancing both the basic techniques and some of the advance techniques in Microbiology, the present revision is related to restructuring of syllabus under autonomy updating it with a few more changes.

With the basic knowledge of microbiology concepts and techniques learnt in FYBSC, for upskilling students they are exposed to different biomolecules like proteins, enzymes and DNA. The advance analytical techniques used for detailed understanding of individual biomolecules have been incorporated in the same units for better correlation of technical understanding. Various applied fields like Food microbiology, Dairy microbiology have also been included. An attempt has been made to introduce students to immunology, diagnostics and biostatistics as well.

Learning fundamental processes of genetics, progressively students are also getting exposed to basic techniques of recombinant DNA technology. Interesting practicals related to these topics would add up to the learner's knowledge through experiential learning.

Eligibility:Cleared F.Y.B.Sc. with Microbiology as one of the subjects

Duration:3 years

Mode of Conduct:Laboratory Practicals/Offline lectures/Online lectures

Program Specific Outcome:

At the end of the program Students would be able to:

- Study and identify various microbial cultures
- Analyse the role in their natural niche.
- Apply knowledge to make positive use of microbial systems for betterment of community and for sustainable development.

VPM's B.N.Bandodkar College of Science (Autonomous), Thane
S.Y.B.Sc.
(Microbiology) Structure of Programme

CourseCode	CourseTitle	No.oflectures	Credits
BNBUSMB3T1	Biomolecules, Bio-analytics and Microbial taxonomy	45	2
BNBUSMB3T2	Applied Environmental Microbiology	45	2
BNBUSMB3T3	Microbial Genetics	45	2
BNBUSMB3P3	Section I (Practicals Based on Units I, II and III of BNBUSMB3T1)	45	1
	Section II (Practicals Based on Units I, II and III of BNBUSMB3T2)	45	1
	Section III (Practicals Based on Units I, II and III of BNBUSMB3T3)	45	1
Total		270	9

CourseCode	CourseTitle	No.oflectures	Credits
BNBUSMB4T1	Bioenergetics, Enzymology and Protein Analysis	45	2
BNBUSMB4T2	Industrial Technology, Food and Dairy Microbiology	45	2
BNBUSMB4T3	Immunology, clinical microbiology and basic biostatistics	45	2
BNBUSMB4P4	Section I (Practicals Based on Units I, II and III of BNBUSMB4T1)	45	1
	Section II (Practicals Based on Units I, II and III of BNBUSMB4T2)	45	1
	Section III (Practicals Based on Units I, II and III of BNBUSMB4T3)	45	1
Total		270	9

Semester III

Course CodeBNBUSM B3T1	CourseTitle Biomolecules,Bio-analyticsandMicrobialtaxonomy	Credits 2	No. oflectur es 45
LearningOutcomes: Learnerwillbeableto <ul style="list-style-type: none"> • Understandstructureandchemistryofnucleic acid • Carryoutestimationofbiomolecules • Performagarosegelelectrophoresisandbasicchromatography • Knowthe significanceofmodern toolsof microbialtaxonomy 			
Unit I:Nucleic acidstructure andchemistry	<p>1.1 NucleicAcidStructure</p> <ul style="list-style-type: none"> a. Properties of DNA b. Structural features of DNA c. Properties of RNA d. Structural features of RNA <p>1.2 Nucleicacidchemistry</p> <ul style="list-style-type: none"> a. Denaturationofdouble helicalDNA andRNA b. Nucleicacidfromdifferentspeciescanformhybrids c. Nucleotidesandnucleicacidsundergononenzymatictra nsformations d. DNAmethylation <p>1.3 OtherFunctionsofnucleotides</p> <p>1.4 DNAsupercoilingand linkingnumber</p> <p>1.5 Structuresofchromosomesofeukaryoticcell</p>		15

Unit II Bio-analytics	<p>Estimation of Biomolecules:</p> <p>2.1 Macromolecular composition of a microbial cell</p> <p>2.2 Methods of elemental analysis: Carbon , Nitrogen and Phosphorus</p> <p>2.3 Quantitative Estimation of Biomolecules</p> <p>Proteins: Biuret, Lowry & Bradford methods (Comparative analysis)</p> <p>Carbohydrates: DNSA method</p> <p>Lipids: Soxhlet method</p> <p>DNA: DPA</p> <p>mRNA: by Orcinol method</p> <p>Basic separation methods:</p> <p>2.4 Chromatography: Principles of Chromatography</p> <ul style="list-style-type: none"> A. Planar Chromatography: Paper Chromatography & Thin-Layer Chromatography B. Column Chromatography <p>Types of Chromatography: (principle & working)</p> <ul style="list-style-type: none"> 1. Adsorption Chromatography 2. Gas-Liquid Chromatography (GLC) 3. Gel Permeation Chromatography 4. Ion Exchange Chromatography 5. Affinity Chromatography 6. High Performance Liquid Chromatography <p>2.5 Agarose Gel Electrophoresis</p> <p>Ninhydrin, Anthrone</p>	15
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Unit III Microbiology taxonomy	<p>3.1 Introduction to Microbial Classification and Taxonomy: Systems of Classification: Phenetic, Phylogenetic, Genotypic, Numerical Taxonomy;</p> <p>3.2 Taxonomic Ranks: Species, Genus, Family, Order, Class, Phylum, Domain</p> <p>3.3 Techniques for determining Microbial taxonomy and Phylogeny: Morphological, physiological, metabolic and ecological characteristics Genetic Analysis, Molecular characteristics: Nucleic acid base composition, Nucleic acid Hybridization, nucleic acid and amino acid sequencing, Fingerprinting</p> <p>3.4 Concept of uncultured bacterial diversity; attempts to culture the 'yet uncultured bacteria': Multilocus sequence analysis, Fatty acid analysis</p>	15
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CourseCode BNBUSMB3T2	CourseTitle Applied Environmental Microbiology	Credits 2	No. oflectur es 45
LearningOutcomes: Learner will be able to			
	<ul style="list-style-type: none"> • Demonstrate the presence of air microflora and relate it to air quality • Illustrate various biogeochemical cycles • Perform water analysis • Justify the use of microbes as biofertilizers and biopesticides 		
Unit I: Air & Soil Microbiology Air Microbiology	<p>1.1 Air microbiology</p> <ul style="list-style-type: none"> a. Important Airborne Pathogens and Important Toxins released by Airborne bacteria b. Aerosols, Nature of Bioaerosols, Aeromicrobiological pathway c. Microbial survival in the air d. Extramural and Intramural aeromicrobiology (Overview) e. Bioaerosol Control f. Air sampling methods and devices <p>1.2 Soil Microbiology</p> <ul style="list-style-type: none"> a. Composition and types of soil, textural triangle b. Types of soil microorganisms and their activities c. Methods of studying soil microorganisms: Sampling, Cultural methods, Physiological methods, Immunological methods, Nucleic acid based methods, Radioisotope techniques d. Biogeochemical cycles: Carbon, Nitrogen, Phosphorus, Sulfur e. Soil Microbiome (Introduction) <p style="color: red;">Iron cycle,</p>	15	

Unit II: Fresh Water & Sewage Microbiology	<p>2.1 Fresh Water Microbiology</p> <ul style="list-style-type: none"> a. Fresh water environments and micro-organisms found in Springs, rivers and streams, Lakes, marshes and bogs and marine environment b. Potable water definition, Purification of raw water c. Bacteriological standards of potable water Maharashtra pollution control board (MPCB), Central pollution control board (CPCB), Bureau of Indian standards (BIS) World health Organization (WHO)) d. Waterborne Infections e. Index organisms: Indicators of faecal pollution f. Bacteriological analysis of water for potability: Presumptive (Multiple tube fermentation -MPN), Confirmed, Completed) 	15
	<p>2.2 Sewage and Waste Water Microbiology</p> <ul style="list-style-type: none"> a. Wastewater analysis: Physico-chemical parameters: pH, temperature, TS, COD, BOD, Toxicity b. Industrial water pollutants c. Ecological effects (Biomagnification and Eutrophication) and health hazards d. Removal of Pathogens by Sewage treatment Processes, Oxidation Ponds and Septic tanks, Sludge Processing, Disposal of treated wastewater and biosolids <p style="color: red;">Waste water treatment, Nature of waste water</p>	

<p>Unit III Biofertiliser, Biopesticide, Bio remediation Soil and Geo Microbiology</p>	<p>3.1 Biofertilizers: Soil & Plant Microbial communities</p> <p>3.2 Microbes as Sustainable Biofertilizers: Need & Advantages of Biofertilizer Over Chemical</p> <p>3.3 Introduction to types of Biofertilizers: Non-symbiotic & Symbiotic Nitrogen fixation, Phosphate Solubilizing Microorganisms, potassium mobilization, Green Manuring, Cyanobacterial inoculants, Azo lla, Frankia induced nodulation, Mycorrhizal Fungi</p> <p>3.4 Methods of application of Biofertilizers</p> <p>3.5 Biopesticides: Introduction; Types of Biological Control (Classical, inoculation, Inundation), Examples each of Bacterial, Viral, Fungal and Protozoal biopesticide, Concept of Biofungicide and Bioherbicide</p> <p>3.6 Bioremediation, Bioaugmentation, Biotransformation, Xenobiotics, Recalcitrant xenobiotics, Biomagnification</p> <p>3.7 Factors affecting bioremediation & Types of Bioremediation (<i>in-situ, ex-situ</i>)</p> <p>3.8 Applications of Bioremediation - hydrocarbons, (Oil spills) Pesticides and herbicides, Heavy metals (Uranium) contaminated soil and wasteland, Ground Water; Genetically modified microbes in bioremediation</p> <p>3.9 Advantages and limitations Mass production, constraints in biofertilizer technology and biofertilizer strains develop Basic requirement for establishment of biopesticide, technical aspect, major biopesticides produced and used in India, Biopesticide formulations</p>	15
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Course Code	Course Title	Credits	No. of Lectures
BNBUSMB3T3	Basic genetics and Recombinant DNA technology	2	45

Learning Outcomes: Learners will be able to:

- Understand the molecular mechanisms of DNA replication
- Differentiate between DNA replication in prokaryotes and eukaryotes
- Explain basic terminologies related to microbial genetics.
- Know the significance and components of basic PCR

Unit I Replication <i>Basics of Microbiology</i>	1.1 Evolutionary importance of DNA as genetic material 1.2 DNA replication in prokaryotes and eukaryotes- 1.3 Semi-conservative DNA replication: the Meselson and Stahl Experiment, 1.4 Role of different proteins and enzymes in DNA replication: Initiator proteins, Helicases, Primase, SSBPs, DNAGyrases, DNA ligase 1.5 Semi-discontinuous Replication: The Okazaki experiment 1.6 Bidirectional Replication of circular DNA molecules, pulse chase experiment 1.7 Molecular model of DNA Replication in <i>E. coli</i> : Initiation, elongation and termination of replication 1.8 DNA polymerases and their role 1.9 Rolling circle replication 1.10 DNA replication in Eukaryotes: Replicons, Initiation of replication, replication enzymes, Replicating the ends of chromosomes, telomerase: action and significance	15
Unit II Transcription <i>Physical and chemical agent for microbial control</i>	<p>2.1 Central Dogma: An Overview, Transcription process, Transcription in bacteria –</p> <ul style="list-style-type: none"> • Initiation of transcription at promoters, • elongation of an RNA chain, • termination of an RNA chain <p>2.2 Transcription in Eukaryotes - Eukaryotic RNA polymerase, 2.3 Transcription of protein- coding genes by RNA polymerase II, Transcription initiation, The structure and production of Eukaryotic mRNAs, Production of mature mRNA in Eukaryotes, Processing of Pre-mRNA to mature mRNA. Self Splicing of Introns, RNA editing</p> <p>2.4 Genetic code - Nature of genetic code and characteristics of genetic code.</p> <p>2.5 classes of RNA and its significance</p>	15
Unit III Basic Recombinant DNA Technology Basic rDNA technology	3.1 Introduction 3.2 Model organisms: prokaryotic, eukaryotic 3.3 Chemical Synthesis of DNA. 3.4 Restriction endonucleases, DNA ligase 3.5 <i>E. coli</i> : preferred host for cloning 3.4 Vector: pBR322, design, advantages and drawbacks 3.5 Polymerase Chain Reaction: General Principle, 3.6 Components of a Typical PCR Reaction; Basic Experimental Design	15

and bioinformatics		
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courseCode	CourseTitle	Credits	No. oflectur es
BNBUSMB3P3	PracticalBasedonBNBUSMB3T1,B NBUSMB3T2&BNBUSMB3T3	3	
	SectionI		
Practical1	PracticalsBasedonUnit IofBNBUSMB3T1		
a.	IsolationofgenomicDNA from <i>E.coli</i>		
Practical2	PracticalsBasedonUnit IIofBNBUSMB3T1		
a.	Estimationof Proteinby Biuretmethod		

b.	Estimation of sugars by DNSA method	
c.	Estimation of DNA by DPA method	
d.	Estimation of RNA by Orcinol method	
e.	Separation of Amino acids by Paper Chromatography	
f.	TLC of sugars	
g.	Demonstration of Size exclusion Chromatography	
h.	Demonstration of AGE	
Practical3	Practicals Based on Unit III of BNBUSMB3T1	
a.	Isolation and identification of bacteria isolated from different ecosystems	
b.	Study of bacterial diversity using dilute cultivation media	
c.	Demonstration : Analysis of phylogenetic tree	
	Section II	
Practical4	Practicals Based on Unit I of BNBUSMB3T1	
a.	Gravity sedimentation method for Air microflora analysis	
b.	Determination of Soil pH and Soil moisture content	
c.	Study of soil ecosystem by contact slide method	
d.	Enrichment and isolation of microorganisms from soil: a. Nitrifying bacteria (in liquid medium) qualitative detection; b. Ureolytic bacteria (Broth and Agar), c. Cellulolytic bacteria (liquid medium qualitative detection) and isolation	
e.	Isolation of Phosphate solubilizing bacteria	
Practical5	Practicals Based on Unit II of BNBUSMB3T2	
a.	Winogradsky's column	
b.	Routine analysis of water: a. Standard Plate Count	

	b. Detection of Coliforms in water: Presumptive Test, Confirmed Test and Completed Test c. Rapid Detection of <i>E. coli</i> by MUG Technique (Demonstration)	
c.	Wastewater analysis: a. Study of microbial flora in raw and treated sewage b. Determination of BOD and COD of wastewater	
Practical6	Practicals Based on Unit III of BNBUSMB3T2	
a.	Isolation of Blue Green Algae from Water samples	

b.	Isolation of <i>Azospirillum</i>	
c.	Isolating Chromium tolerant organisms	
Section III		
Practical 7	Practicals Based on Unit I and II of BNBUSMB3T3	
a.	Reagent preparation for molecular biology techniques	
Practical 8	Practicals Based on Unit III of BNBUSMB3T3	
a.	Restriction digestion of lambda phage/any plasmid DNA	
b.	Ligation reaction using <i>EcoRI</i> digested λ DNA fragment	
c.	Isolation and visualization of plasmid DNA	
	Estimation of total sugar by Anthrone method (Demo), Estimation of reducing method by Fehling's method, Extraction of lipid by Soxhlet method, Enumeration of microorganisms in air and study of its load after fumigation, Study of air microflora and determination of sedimentation rate, Determination of total solids in wastewater	

SemesterIV

CourseCode BNBUSMB4T1	CourseTitle Bioenergetics,EnzymologyandProteinAnalysis	Credits 2	No. oflectur es 45
LearningOutcomes: Learners will be able to:			
	<ul style="list-style-type: none"> Understand basic aspects of bioenergetics and metabolism Comprehend the basic principles of enzymology Enlist the mechanisms underlying the enzymatic reactions and Know different methods used for protein purification and separation 		
Unit I:Introduction toMetabolism &Bioenergetics	1.1 Introduction to metabolism, Metabolic pathway 1.2 Organic reaction mechanisms 1.3 Experimental approaches to study metabolism 1.4 Thermodynamics of Phosphate compounds 1.5 Oxidation-reduction reactions 1.6 Thermodynamics of life		15
Unit II:EnzymeKineti cs	2.1 History, Definition, Classification and Nomenclature of enzymes 2.2 Chemical Nature, Properties of Enzymes, 2.3 Mechanism of Enzyme Action, 2.4 Michaelis-Menten equation and its derivation; Lineweaver-Burk plot 2.5 Overview of Coenzyme: a. Coenzymes: Different types and reactions catalyzed by coenzymes (in tabular form) b. Nicotinic acid: structure, occurrence & biochemical function 2.6 Enzyme Kinetics: a. Saturation kinetics b. Effect of temperature and pH c. Effect of Inhibitors- Reversible and irreversible, competitive, Non-competitive and uncompetitive inhibitors 2.7 Multi-substrate reactions- Ordered, Random and ping pong reactions 2.8 Allosteric effects in enzyme catalyzed reactions- Koshland-Nemethy and Filmer model & Monod, Wyman and Changeux Model		15

<p>Unit III:Protein Analysis Analytical techniques</p>	<p>3.1 Purificationofproteins: a. Recognizingthe proteinof interest b. Criteria for protein purification: Solubility, Size,Charge, and Binding Affinity 3.2 Electrophoresisofproteins: a. SDS-PAGE</p>	<p>15</p>
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	<p>b. NativeGels, Gradientgels, Isoelectricfocusinggels c. 2DPAGE d. Celluloseacetateelectrophoresis e. Detection, estimation, recoveryofproteinsin gels</p> <p>3.3 Basicprinciple ofsedimentation:</p> <ul style="list-style-type: none"> a. RCFandsedimentationconcepts b. Sedimentationrateorvelocity c. Sedimentationequilibrium d. Svedbergunits <p>3.4 Construction,workingofcentrifugetypes:</p> <ul style="list-style-type: none"> a. Typesofcentrifuges b. Typesofrotors c. Care andmaintenance ofcentrifuges <p>3.5 Preparativecentrifugation:</p> <ul style="list-style-type: none"> a. Differentialcentrifugation, b. Density gradient <p>centrifugationApplicationsofpreparatorycentrifuge</p>	
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Course CodeBNBUSM B4T2	CourseTitle IndustrialTechnology,FoodandDairyMicrobiology	Credits 2	No. oflectur es45
LearningOutcomes: Learnerswillbeableto:			
	<ul style="list-style-type: none"> • Namedifferentcomponentsofbasicfermenterwiththeirfunctionalsignificance • Comparedifferentmethodsoffoodpreservations • Demonstratethepresenceofdifferentspoilagecausingorganism • Explainvariousmethodusedinanalysisofmilk 		
Unit IIIndustrialTe chnology Host defense and public health	<p>1.1 Anintroductiontofermentationprocesses</p> <p>1.2 Therangeoffermentationprocesses</p> <p>1.3 TheComponent partsofafermentation process</p> <p>1.4 Screeningmethods:PrimaryandsecondaryscreeningHighthroughput screening methods</p> <p>1.5 Strain improvement: selection of induced mutants synthesizingimproved levels of primary metabolites; The isolation ofinduced mutants producing improved yields of secondarymetabolites; The improvement of strains by modifyingpropertiesother than theyield ofproduct</p> <p>1.6PreservationofculturesandQualitycontrolofpreservedstock</p> <p>1.7 Types of fermentations: Surface and Submerged; Batch andContinuous,Aerobic andanaerobic</p>		15
Unit IIFood Microbiology	<p>2.1Microbesasfood, overviewoffermentedfoods</p> <p>2.2.Microbial growth andfood spoilage</p> <p>2.3 Controllingfoodspoilage:</p> <ol style="list-style-type: none"> a. Homeostasisandhurdletechnology b. Naturally occurring antimicrobials: lysozyme, lactoferrin andother Fe binding proteins, avidin, spices and essential oils, onions and garlic, isothiocyanates, phenolic compounds c. Antimicrobial chemicals: organic acids, nitrites, Para-benzoicacid,sodium chloride, phosphates,sulfites d. Biopreservation:controlledacidification,bacteriocinsprobiotics,prebiotics andsymbiotics <p>2.4 Physicalmethodsoffoodpreservation:drying,freeze-drying,coldstorage,heat treatment:concept of TDPandTDT</p> <p>2.5 Preservationbyirradiation:UVandionizingradiations</p>		15

	2.6 Food Control Enforcement & Control Agency: International agencies, Federal agencies (FDA, USDA), FSSAI [website], Introduction to HACCP Important Microorganisms in Food Microbiology: Spoilage causing microorganisms, Food-borne Illness associated Microorganisms	
Unit III Dairy Microbiology	3.1 Microbiology of raw milk, spoilage causing organisms, sources of contamination of milk, important pathogenic microbes 3.2 Microbial quality of raw milk 3.3 Milk analysis: Grading, platform tests, Dye reduction test, DMC, SPC, LPC, Coliform count, Thermophilic count, Psychrophilic count 3.4 Rapid detection of milk borne pathogens (Nucleic acid based assays, Immunological based assays, Biosensors) 3.5 Products: Cheese, Butter, Yoghurt (types, manufacturing process outline, Defects) 3.6 Microbiological Quality and Safety Aspects of Traditional Dairy Products	15

CourseCode BNBUSMB4T3	CourseTitle Immunology, clinical microbiology and basic biostatistics	Credits 2	No. of lecturers 45
Learning Outcomes: Learners will be able to:			
Unit I: Immunology Nanotechnology, Biofilms and biosensors with application	<ul style="list-style-type: none"> • Explain role of lymphoid cells, organs, immunoglobulin • Explain conventional, rapid and advance methods used in isolation and identification bacteria from clinical sample • Represent data with statistical tools <p>1.1 Classification of immune components of innate immunity & acquired immunity 1.2 Cells of immune system: Hematopoiesis, lymphocytes, monocytes & macrophages, granulocytes, mast cells, dendritic cells & NK cells 1.3 Lymphoid organs : primary and secondary 1.4 Concept of local immunity and herd immunity 1.5 Basic concept of antigen and epitope, types of antigen 1.6 Basic structure, classes and function of immunoglobulin</p>		15

UnitII: ClinicalMicrobiology Scientific writing, research methodology and biostatistics	2.2 PhasesofDiagnosticcycle 2.3 Clinicalmicrobiology:SpecimenCollectionand handling 2.3 ApproachestoPathogen identification ConventionalApproach 2.4 RapidTechniquesfor pathogenidentification 2.5 Otherpathogensidentifiedbyconventionalorrapiddiagnostics 2.6 Point-of-careRapid Diagnostics	15
Unit III: Basicbiostatistics Biofertiliser, biopesticide, bioremediation	<p>3.1 Introductiontobiostatistics:</p> <ul style="list-style-type: none"> a. Statistics,Biostatistics,Applicationsandusesof Biostatistics b. Data: Types of data (Qualitative/Quantitative,Primary/Secondary,Univariable/Bivariable/Multivariable.Rawdata/Derived data) c. Population,Sampling,Sample,Randomsample,Characteristic(Measurable/ Non-measurable) <p>3.2 Frequencydistribution</p> <ul style="list-style-type: none"> a. Typesoffrequencydistribution(Simple/Grouped) b. Terms associated with grouped frequency distribution (Classinterval, class limit, class boundaries, class mark, class width,classfrequency,cumulativefrequency) <p>3.3 Graphicalrepresentationofdata:</p> <ul style="list-style-type: none"> a. Graphicalrepresentationofungroupeddata:Linegraph,Bardigram(simple), piechart b. Graphical representation of grouped data: histogram (equalclassintervals), frequency polygon <p>3.4 Centraltendency:</p> <ul style="list-style-type: none"> a. Characteristicsofcentraltendency b. Measures of central tendency: mean, median, mode(Ungroupeddata, grouped data) <p>3.5 Measuresofvariation: Absoluteandrelativemeasureofdispersion Forgroupedandungroupeddata:Standard deviation</p> <p>3.6 Significance of Statistics in Epidemiology</p> <p>3.7 Epidemiological techniques and markers</p>	15

CourseCode BNBUSMB4P4	CourseTitle Practicals Based on BNBUSMB4T1,BNBUSMB4T2& BNBUSMB4T3	Credits 3	No. oflectu res
	SectionI		
Practical1	PracticalsBasedonUnit IofBNBUSMB4T1		
a.	Problemsbasedonbioenergetics to calculate K_{eq} , Gibbs energy, Enthalpy		
b.	Radioisotopes half-life calculations		
Practical2	PracticalsBasedonUnit IIofBNBUSMB4T1		
a.	Effect of temperature, pH, enzyme concentration, substrate concentration and inhibitors on Invertase activity		
Practical3	PracticalsBasedonUnit IIIofBNBUSMB4T1		
a.	Protein precipitation using ammonium sulphate		
b.	Desalting using dialysis method		
c.	Demonstration of Affinity chromatography		
d.	Demonstration of Ion exchange chromatography		
e.	Electrophoresis of proteins by SDS-PAGE		
f.	Isolation of chloroplast using sucrose density gradient centrifugation		
	SectionII		
Practical4	PracticalsBasedonUnit IofBNBUSMB4T2		
a.	Production of amylase by solid state and submerged fermentation		
Practical5	PracticalsBasedonUnit IIofBNBUSMB4T2		
a.	Isolation of organisms causing Food Spoilage		
b.	Determination of MIC of salt/sugar		
c.	Determination of TDP and TDT		
Practical6	PracticalsBasedonUnit IIIofBNBUSMB4T2		
a.	Dye reduction test: 1) MBRT 2) RRT		
b.	Microbiological Quality Analysis of local dairy product 1. Coliform count 2. Thermophilic count 3) Psychrophilic count 4) Fecal streptococcal count 5) Yeast and mold count		
	SectionIII		
Practical7	PracticalsBased onUnit IofBNBUSMB4T3		

a.	Separation of PBMNCs using Ficoll Hypaque	
b.	Field's staining of Blood film	
Practical8	Practicals Based on Unit II of BN BUS MB4T3	
a.	Isolation of organisms on XLD, CLED and Cetrimide agar	
b.	<p>Biochemical test:</p> <ul style="list-style-type: none"> a. Triple sugar iron slant b. Catalase c. Oxidase d. Lysine decarboxylase e. Oxidative-Fermentative Test 	
c.	Demonstration of PCR	
Practical9	Practicals Based on Unit III of BN BUS MB4T3	
a.	Representing data obtained from a self study statistically on graph paper and using MS-Excel: Line graph, Bar diagram, Pie chart	
b.	Solving problems based on measures of central tendency using MS-Excel.	
c.	Solving problems based on measures of dispersion using MS-Excel.	
	Isolation of amylase, protease, lipase producers, Extracellular production of invertase from yeast, Sizing Yeast cells, centrifuge machine	

References:SE

MESTER-III

BNBUSMB3T1:Biomolecules,BioanalyticsandMicrobial taxonomy

Books and References:					
Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Lehninger Principles of Biochemistry	Nelson and Cox	WH Freeman	4 th	2004
2.	Laboratory Manual in Biochemistry	J. Jayaraman	New Age International Publishers		2003
3.	A handbook of Organic Analysis: qualitative and quantitative	Hans Thacher Clarke	CBS publishers & distributors, New Delhi	4 th	2007
4.	Biochemical Methods	S. Sadashivam & A. Manickam	New Age International Publishers	3 rd	2018
5.	Biochemistry Laboratory: Modern Theory & Techniques	Rodney Boyer	Pearson Education, Inc	2 nd	2012
6.	Molecular Cloning	Sambrook & Russel	Cold Spring Harbor Laboratory Press	3 rd	2001
7.	Prescott, Harley & Klein's Microbiology	Willey, Sherwood & Woolverton	McGraw-Hill	7 th	2008
8.	Microbiology - An Evolving science	Joan L. Slonczewski and John W. Foster	W.W. Norton Company, NY	4 th	2017

BNBUSMB3T2:Applied Environmental Microbiology

Books and References:					
Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Environmental Microbiology	Raina M. Maier, Ian L. Pepper & Charles P. Gerba	Academic Press	2 nd	2010
2.	Introduction to Environmental Microbiology	Barbara Kolwzan, Waldemar Adamik	Oficyna Wydawnicza Politechniki Wrocławskiej, Wrocław	-	-
3.	Soil Microbiology	N. S. Subba Rao	Oxford and IBH Publishing Co. Pvt Ltd	4 th	2000

Books and References:

Sr.No.	Title	Author/s	Publisher	Edition	Year
4.	Microbiology- Anevolving science	JoanL.Slonczewskia ndJohnW.Foster	W.WNortonCompany, NY	4 th	
5.	Prescott, Harley & Klein's Microbiology	Willey, Sherwood & Woolverton	McGraw-Hill	7 th	2008
6.	Fundamentals of Microbiology	Frobisher, Hinsdill, Crabtree, Goodheart	SaundersCollege Publishing	9 th	1974
7.	Biotechnology	BD Singh	KalyaniPublishers	4 th	2010
8.	A textbook of Biotechnology	RCDubey	S.Chand	4 th	2006
9.	https://blog.bitsathy.ac.in/an-overview-of-biofertilizers-in-sustainable-agriculture/				
10.	https://www.researchgate.net/publication/335065992_Microbial_Biofertilizers_Types_and_Application				
11.	https://www.researchgate.net/publication/335065992_Microbial_Biofertilizers_Types_and_Application				

BNBUSMB3T3: Basic genetics and Recombinant DNA technology**Books and References:**

Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Molecular biology	DavidFreifelder	Jones&Bartlett Publishers	2 nd	1987
2.	iGenetics: A Molecular Approach	PeterRussel	Benjamin Cummings	3 rd	2010
3.	Molecular Biology	RobertWeaver	McGrawHillinternational edition.	3 rd	2005
4.	Molecular Cloning A laboratory manual (Volume 1)	Sambrook, Russell	CSDL press	3rd	2001

SEMESTER-IV

BNBUSMB4T1:Bioenergetics, EnzymologyandProteinAnalysis

Books and References:					
Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Fundamentals of Biochemistry	D.Voet and J. Voet	Publisher Wileyplus	5 th	2011
2.	Biochemistry	Satyanarayana and Chakrapani	Books and allied pvt ltd.	3 rd	2006
3.	Lehninger Principles of Biochemistry	Nelson and Cox	WH Freeman	4 th	2004
4.	Outlines of Biochemistry	E.E. Conn & P.K. Stumpf, G. Bruening, R.N. Doi.	John Wiley and sons	5 th	1999
5.	Biochemistry	Lubert Stryer	W.H. Freeman and Company	5 th	2002
6.	Principle and techniques of Biochemistry and Molecular Biology	Wilson and Walkar	Cambridge University Press	7 th	2010

BNBUSMB4T2:Industrial Technology, Food and Dairy Microbiology

Books and References:					
Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Industrial Microbiology	Casida L.E.,	New Age International Ltd, Publishers, New Delhi.	Reprint	2009
2.	Principles of Fermentation Technology	Stanbury P. F., Whitaker A. & Hall S.J.,	Aditya Books Pvt. Ltd, New Delhi	2 nd	1997
3.	Dairy microbiology	K.C. Mahanta.	Omson Publication	1 st	1984
4.	Food Microbiology	Adam and Moss	New Age International Ltd.	3 rd	2008
5.	Milk and milk products	Eckles	Tata McGraw Hill Publishing Company Ltd.	4 th	1986
6.	Outlines of dairy technology	Sukumar De	Oxford University Press	13 th	1980

BNBUSMB4T3: Immunology, Clinical Microbiology and biostatistics

Books and References:					
Sr.No.	Title	Author/s	Publisher	Edition	Year
1.	Immunology essential and fundamental	S. Pathak, U Palan	Capital Publishing Company	2 nd	2005
2.	Immunology	Kindt, Goldsby, Osborne	W. H. Freeman and Co.	6th	2007
3.	Microbiology: An Evolving Science	Slonczewski and Foster	Norton & Company, Inc	4 th	2016
4.	Koneman's color atlas and textbook of diagnostic microbiology	Elmer W. Koneman	Lippincott Williams & Wilkins	6 th	2006
5.	Introduction to Biostatistics	Dr. Pranab Kumar Banerjee	S. Chand Publishing	Revised edition	2007
6.	Microbiology	Michael J. Pelczar, C.S. Chan, , Noel R. Krieg	TATA McGraw - Hill	5 th	1995

Evaluation Scheme

Internals:

Classtest	Assignment/ Study tour with report/Journal Movie club presentation/Presentation of mini-research project work/volunteering for Department fest/postermaking/exhibition/ Departmental contribution/ case study presentation/Review writing	Attendance, Active Participation and Leadership Qualities	Total
20M	10 M	10M	40M

Internal Examination: Based on Unit 1 / Unit 2 / Unit 3

Duration: 40 mins

Total

Marks: 20

No. of Questions: 15

Q.1	Answer the following choosing the correct alternative.					10
1	Based on Unit I / II / III					
a		b	c	D		
2	Based on Unit I / II / III					
a		b	c	D		
3	Based on Unit I / II / III					
a		b	c	D		
4	Based on Unit I / II / III					
a		b	c	D		
5	Based on Unit I / II / III					
a		b	c	D		
6	Based on Unit I / II / III					
a		b	c	D		
7	Based on Unit I / II / III					
a		b	c	D		
8	Based on Unit I / II / III					
a		b	c	D		
9	Based on Unit I / II / III					
a		b	c	D		
10	Based on Unit I / II / III					
a		b	c	D		

Q.2 | Answer the following choosing the correct alternative.

10

	1	Based on Unit I/II/III						
	a	b	c	D				
	2	Based on Unit I/II/III						
	a	b	c	D				
	3	Based on Unit I/II/III						
	a	b	c	D				
	4	Based on Unit I/II/III						
	a	b	c	D				
	5	Based on Unit I/II/III						
	a	b	c	D				

Theory Examination: Suggested Format of Question paper

Duration: 2 Hours

Total Marks: 60

- All questions are compulsory

Q.1	Answer any two of the following						16
	a	Based on Unit I					
	b	Based on Unit I					
	c	Based on Unit I					
Q.2	Answer any two of the following						16
	a	Based on Unit II					
	b	Based on Unit II					
	c	Based on Unit II					
Q.3	Answer any two of the following						16
	a	Based on Unit III					
	b	Based on Unit III					
	c	Based on Unit III					
Q.4	Answer any six of the following						06
	a	Based on Unit I/II/III					
	b	Based on Unit I/II/III					

	c	BasedonUnitI/II/III	
	d	BasedonUnitI/II/III	
	e	BasedonUnitI/II/III	
	f	BasedonUnitI/II/III	
	g	BasedonUnitI/II/III	
	h	BasedonUnitI/II/III	
	i	Basedon UnitI/II/III	
	Answer <i>any two</i> of the following		06
	a	BasedonUnitI/II/III	
	b	BasedonUnitI/II/III	
	c	BasedonUnitI/II/III	

Marks Distribution and Passing Criterion for Each Semester

Theory					Practical		
CourseCode	Internal	Min marks for passing	Theory Examination	Min marks for passing	CourseCode	Practical Examination	Min marks for passing
BNBUSMB3T1	40	16	60	24	BNBUSMB3P3	150	60
BNBUSMB3T2	40	16	60	24			
BNBUSMB3T3	40	16	60	24			

Theory					Practical		
CourseCode	Internal	Min marks for passing	Theory Examination	Min marks for passing	CourseCode	Practical Examination	Min marks for passing
BNBUSMB4T1	40	16	60	24	BNBUSMB4P4	150	60
BNBUSMB4T2	40	16	60	24			
BNBUSMB4T3	40	16	60	24			

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